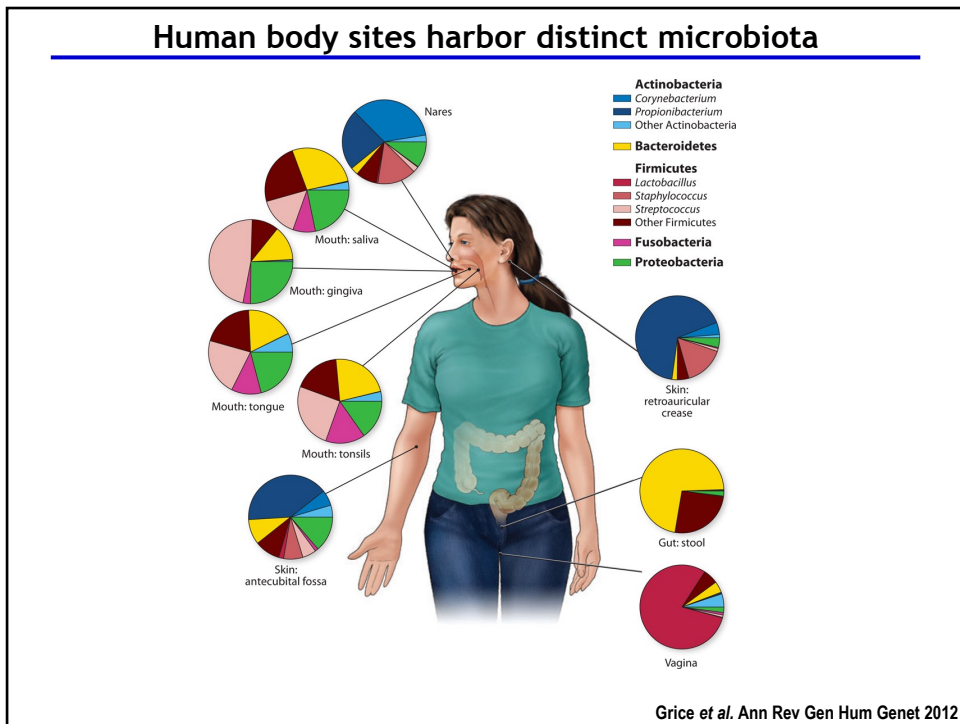


LU3SV649 Microbiota ecology and host immunity

Martin LARSEN

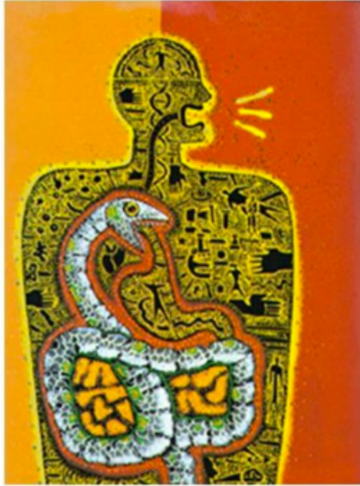
INSERM U1135, CHU Pitié-Salpêtrière, Paris, France

1



10

The human Gut and its inhabitants in numbers



- 30 tons of food and 50.000 L during a lifetime
- Huge mucosal surface: 30-40 m²
- >50 billions of new bacteria every day
- 70-80% of all immune cells are located in the Gut.
- 1-2g secretory IgA per day
- 100 millions of neurons (as many as in the spinal cord).
- 5x10¹³ bacteria: 3x number of cells in the entire body, i.e. 1-2 kg.
- 100 times more bacterial genes than human genes.

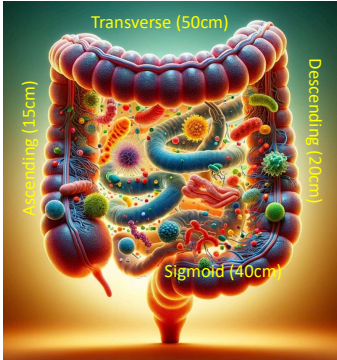
11

The digestive system - A trip through the GI tract

Small intestine

(duodenum -> jejunum -> ileum)

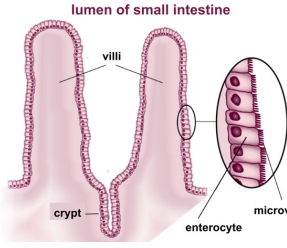
- 3-4m long
- 2.5cm diameter
- Has folds (2x) and villi (8x)
- Has microvilli (8x)
- Surface: 30-40m²
- pH < 7
- Some oxygen
- Host low diversity microbiota.
- Retrieves nutrients



Large intestine

(cecum -> colon -> rectum -> anus)

- 1.5m long
- 4.8cm diameter
- Has no folds nor villi
- Has microvilli (10x surface)
- Surface: 2m²
- pH = 7
- Anaerobic
- Host diverse microbiota.
- Retrieves water and salts.
- Degrades complex nutrients



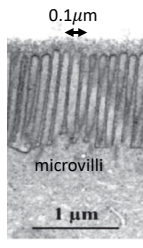
lumen of small intestine

villi

crypt

enterocyte

microvilli

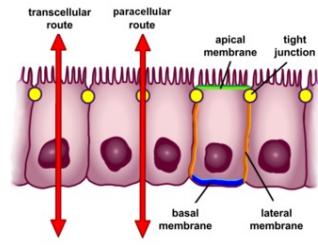


0.1µm

1 µm

microvilli

1 µm



transcellular route

paracellular route

apical membrane

tight junction

basal membrane

lateral membrane

Helander et al. Sc J Gastro 2014

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The digestive system - Anatomy revisited

Small intestine:

Textbook

- Length: 22m
- Diameter: 2.5cm
- Surface: 150-200m² (Tennis court)

Revisited:

- Length: 3-4m
- Diameter: 2.5cm
- Surface 30-40m² (half a badminton court)

Big problems with the measures, because you can mechanically stretch the intestine during e.g. an autopsy.

Helander et al. Sc J Gastro 2014

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Spatial distribution of gut microbiota

> 500 differentes species 10^{14} micro-organisms

Lactobacilli	10^2 to 10^3
Streptococci Lactobacilli	$< 10^{4-5}$
Enterbacteria Enterococcus Faecalis Bacteroides Bifidobacteria Peptococcus Peptostreptococcus Ruminococcus Clostridia Lactobacilli	10^3 to 10^7
	10^9 to 10^{12}

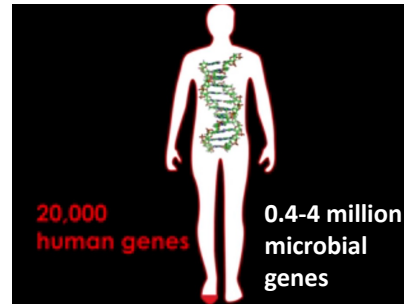
pH

http://www.jpp.krakow.pl/journal/archive/08_15/articles/02_article.html

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We are outnumbered and outsmarted

Outsmarted
(20-200 fold more genes)



Each bacterial species has its own genomic DNA (all human cells have almost identical genomic DNA).

Bacterial genome ($2-10 \times 10^6$ bps) versus human genome (3×10^9 bps)

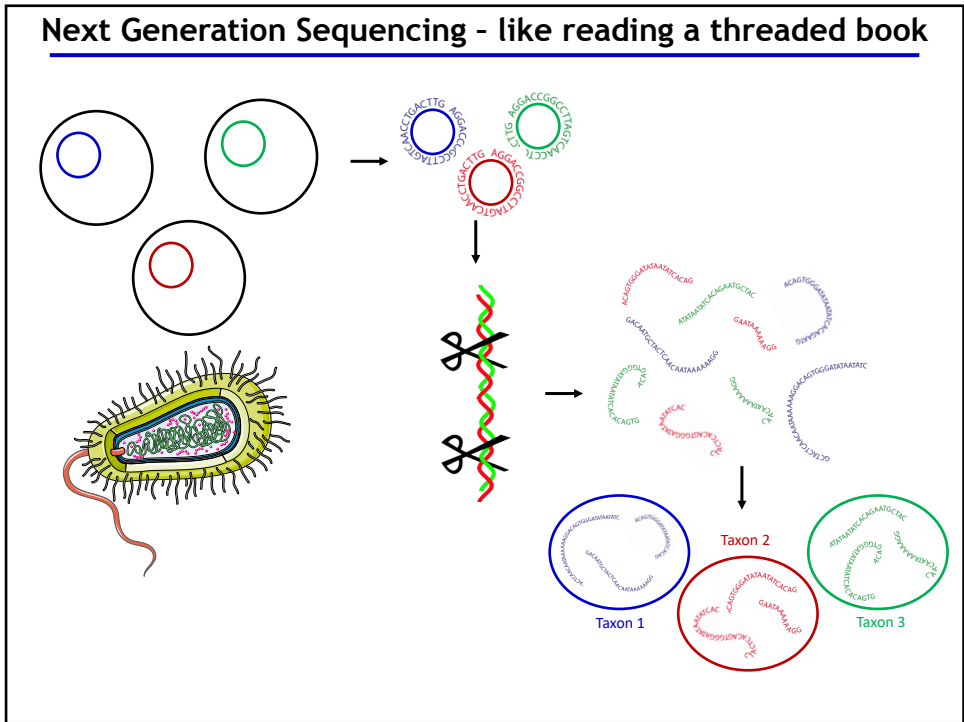
Bacterial DNA is much more compact than human DNA in terms of gene content.

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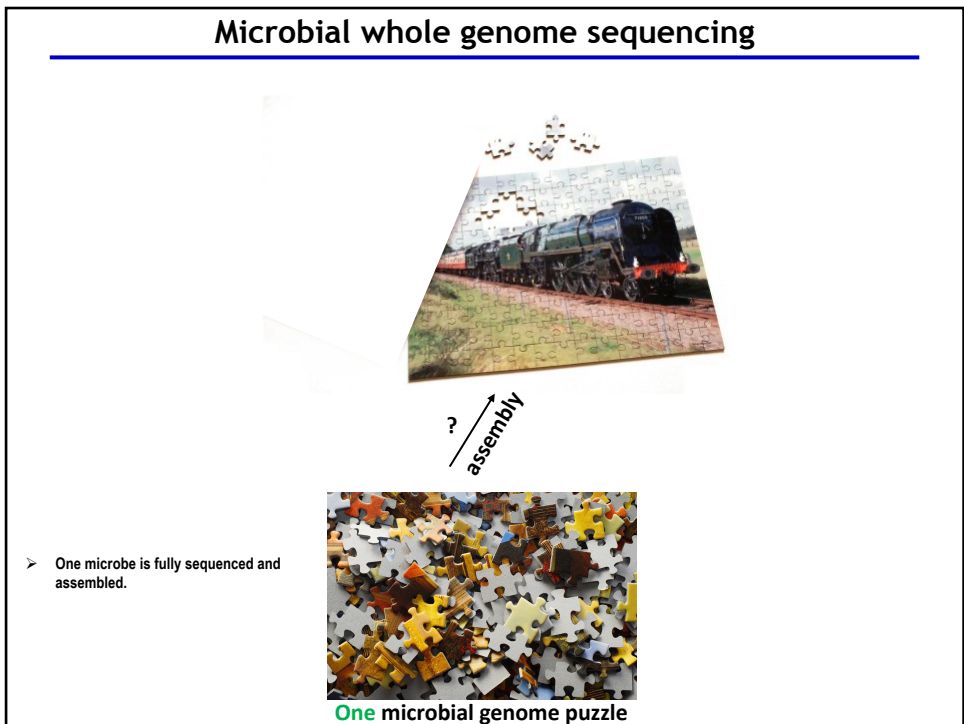
How to study the gut microbiota

- Culture dependent (classic microbiology – aerobiosis/anaerobiosis, medium)
 - Microscopy
 - Metabolism of substrates
 - Co-culture with host cells (epithelium, immune cells etc.)
 - Genetic modifications
 - Culturomics – strain isolation (e.g. FACS)
 - Antibody titers to microbes (ELISA, flow cytometry)
- Culture independent
 - Mass spectrometry
 - Flow cytometry (FISH, immuno-microbiota)
 - Metabolomics
 - Metagenomics (microbiome and immuno-microbiome)
 - Metatranscriptomics

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Microbial whole genome sequencing

The diagram illustrates the process of microbial whole genome sequencing. At the top, a red box labeled "One microbe" contains a puzzle of a train. Below it, a large pile of mixed-colored puzzle pieces is labeled "One microbial genome puzzle". A blue arrow labeled "mapping" points from the pile of pieces to the train puzzle, indicating that individual microbial genomes are mapped back to a reference genome.

➤ Using the reference lid jigsaw genes derived from the reference may be identified.

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Metagenomic sequencing

The diagram illustrates metagenomic sequencing. At the bottom center, a pile of mixed-colored puzzle pieces is labeled "Mixture of microbial genomes". Arrows with question marks point from this pile to four different puzzle images: Mickey Mouse, a train, the Taj Mahal, and a yellow duck. This represents the process of identifying individual microbial genomes from a complex mixture.

➤ A mixture of many microbes

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Metagenomic sequencing

➤ A mixture of many microbes can only be identified if sufficiently many microbial genomes have been fully sequenced.

➤ Alternative – Metagenomic species

Mixture of microbial genomes

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16S rRNA sequencing

A = Acceptor site
P = Peptidyl site
E = Exit site

- 16S rRNA in orange
- Associated protein subunits in blue

Protein synthesis in the

1. Cytosol
2. Endoplasmic reticulum

ER-docking via signal-recognition particle

16S rRNA primary and secondary structures

0 100 200 300 400 500 600 700 800 900 1000 1100 1200 1300 1400 1500 bp

V1	V2	V3	V4	V5	V6	V7	V8	V9
----	----	----	----	----	----	----	----	----

CONSERVED REGIONS: unspecific applications
VARIABLE REGIONS: group or species-specific applications

16S RNA

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16S rRNA sequencing

assembly

Mixture of microbial 16S rRNA genes

- 16s RNA gene is highly variable and functional as a phylogenetic marker.
- Limitation – species from same genus may be undistinguishable (blue sky jigsaws may be hard to distinguish).

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Big data in Microbiology

Intestinal tract

Genomic DNA

Next Generation Sequencing

Identification:

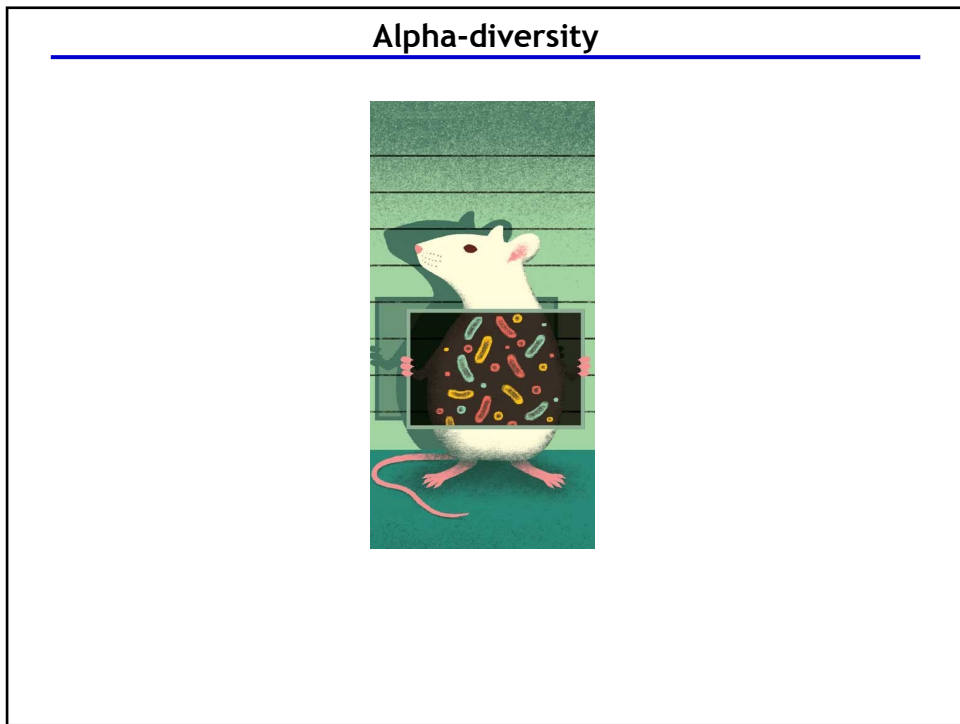
1. Ref. genome
2. Co-abundance clustering

MetaGenomic Species (MGS)

16S rRNA: Pruesse et al. NAR 2007 (www.arb-silva.de)
 Metagenomics: Qin et al. Nature 2010

Nielsen et al. Nat Biotech 2014

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Alpha-diversity

Quantitative
Biomass (g)

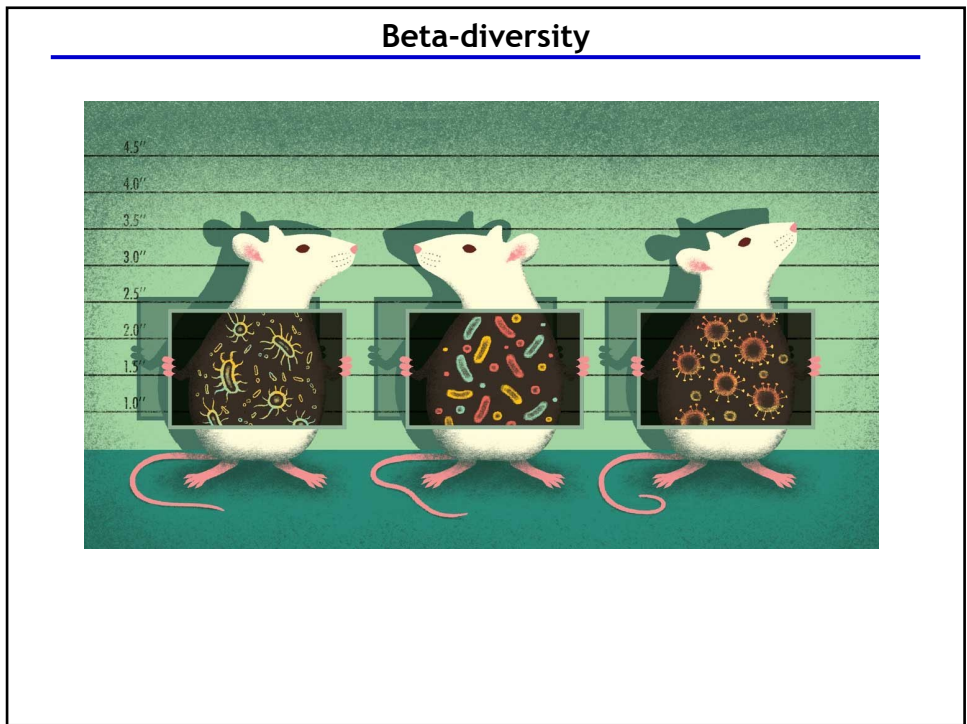
- A community's biodiversity correlates with its size and location
- Ecologists measure biodiversity as heterogeneity, which considers both diversity factors: richness and relative abundance.
- $H = -\sum_{i=1}^k p_i \log(p_i)$; p=abundance
- $E_H = \frac{H}{\log(k)}$; [Normalized]

Qualitative
Richness
Diversity

Species richness	5	5	2	1
Shannon entropy [normalized]	$p_i=1/5$ $H=\log(5)$ $[E_H=1]$	$p_{red} \sim 1 p_{NOT red} \sim 0$ $H \sim -0$ $[E_H \sim -0]$	$p_i=1/2$ $H=\log(2)$ $[E_H=1]$	$p_i=1$ $H=0$ $[E_H=0]$

- Scores of alpha-diversity: Richness, Shannon, Simpson, Chao1 and Chao2
- Chao1 and Chao2 often used in microbiota research. Used when certain species are rare (based on expected rather than observed number of species in a sample).

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Beta-diversity

- Total species diversity (γ) is determined by the mean species diversity of a habitat (α) plus the differentiation among habitats (β). **Robert Whittaker, 1960**
- Pair-wise beta-diversity is measured as similarity or dissimilarity between two samples.
- Beta-diversity is also referred to as species turn-over.
- Absolute species turn-over: $\beta = (R_1 - C_{12}) + (R_2 - C_{12})$, R=Richness, c=common species

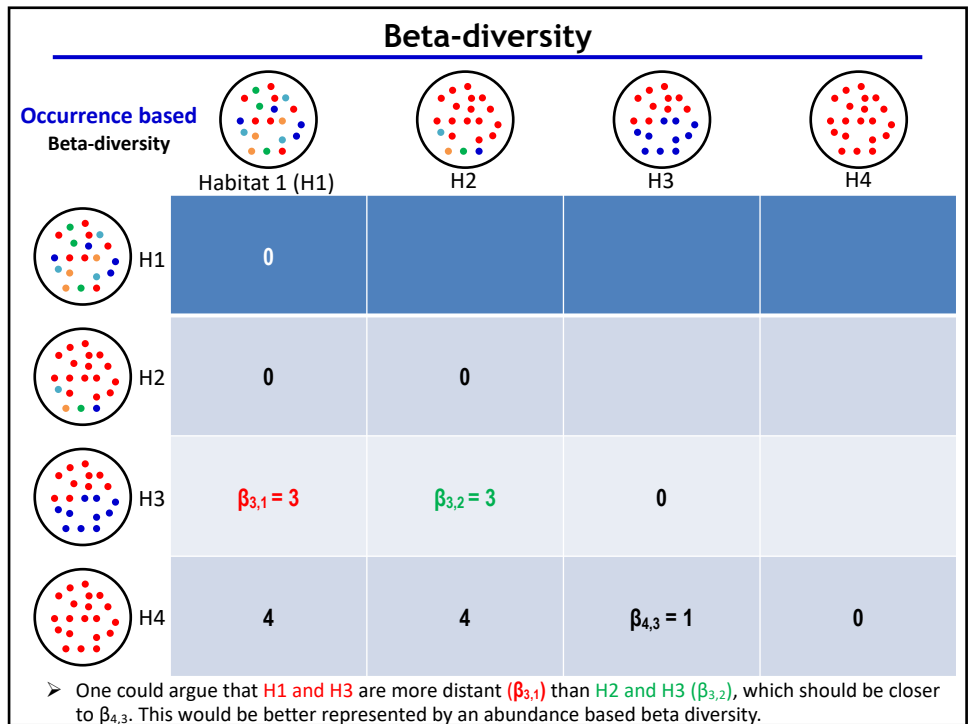
$\beta = (5-5) + (5-5) = 0$

$\beta = (5-2) + (2-2) = 3$ unshared species

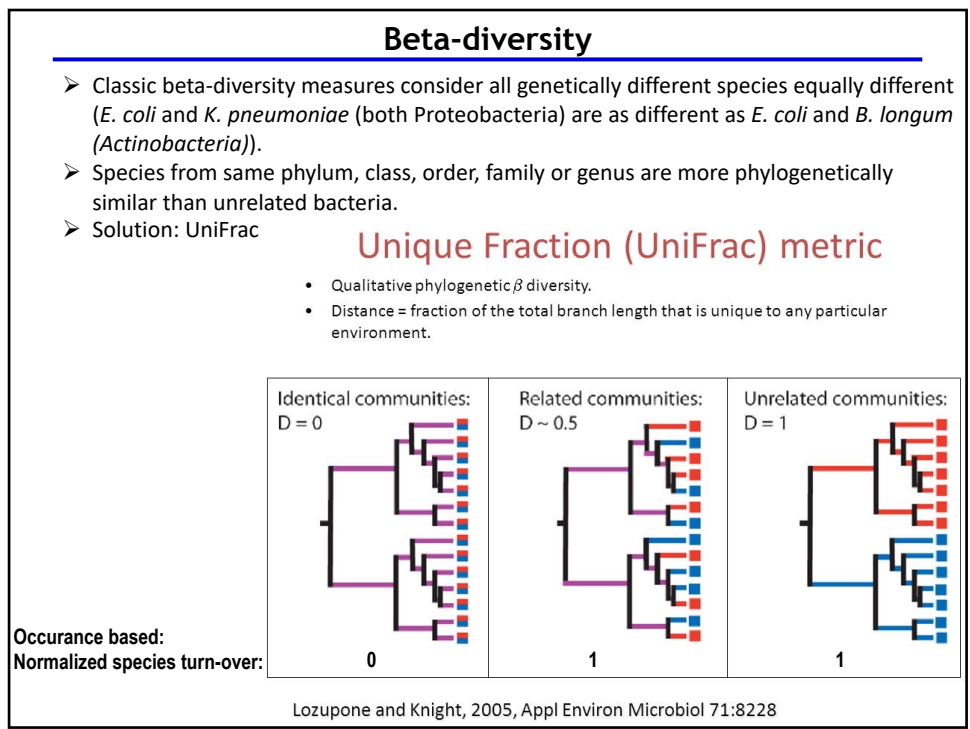
Species richness	5	5	2	1

- Absolute species turn-over is problematic when working with rare species, whose occurrence is associated with random sampling efficiency. Abundance based measures are therefore more appropriate for microbiota work.
- Scores of beta-diversity:
 - Occurrence based: Absolute species turn-over, Whittaker species turn-over (special case of Sørensen similarity index) and Proportional species turn-over (Jaccard similarity index).
 - Abundance based: Bray–Curtis dissimilarity index, Morisita-Horn overlap index.

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Early-life microbiota colonization

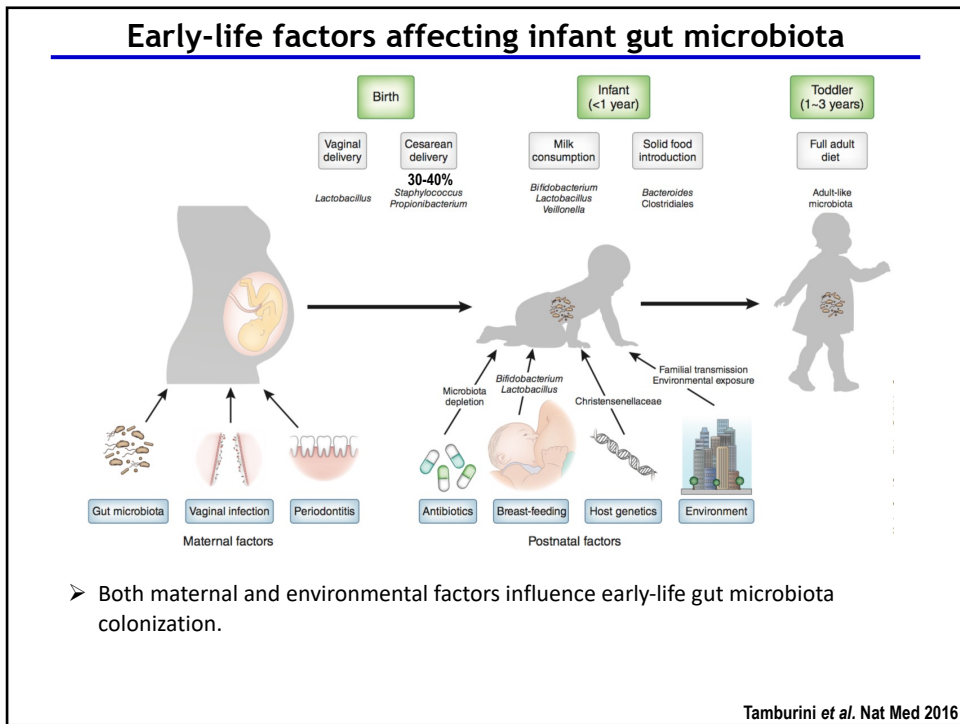
40

Gut microbiota maturation during first 2 years of life

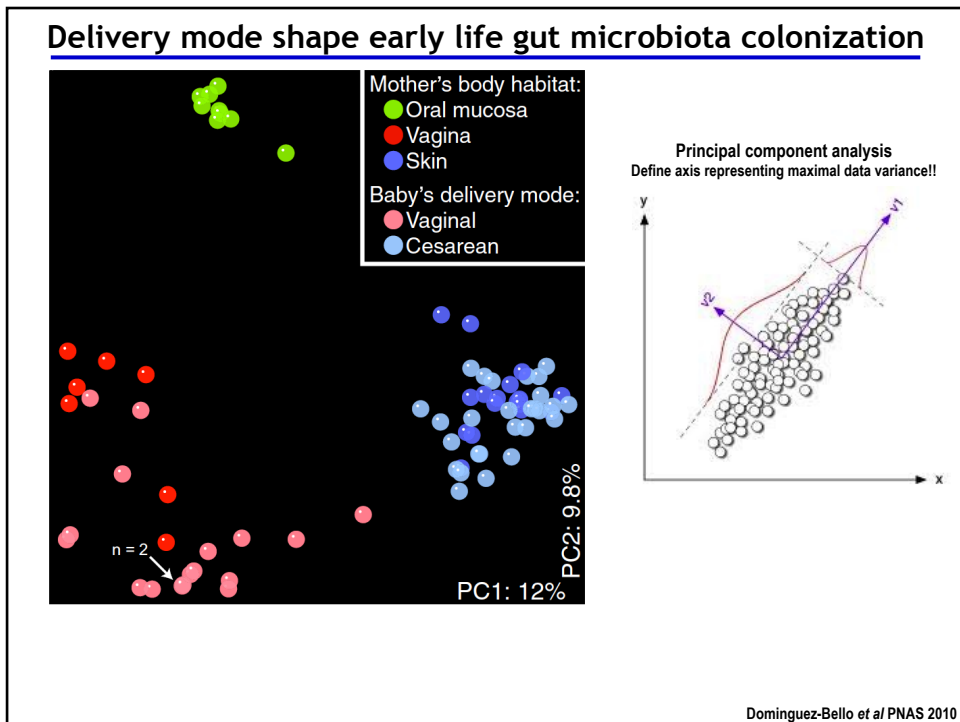
How to avoid deleterious effect of antibiotics on gut microbiota, while preserving systemic anti-microbial activity?

Watch video: <http://www.immulab.fr/cms/index.php/teaching/immunology-teaching> Dr. Rob Knight

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45

Fecal Microbial Transfer (FMT) rescue microbiota post c-section.

Article

Cell

Maternal Fecal Microbiota Transplantation in Cesarean-Born Infants Rapidly Restores Normal Gut Microbial Development: A Proof-of-Concept Study

- Fecal microbiota development of newborns is dependent on the mode of delivery
- The development in cesarean section-born infants deviates from that of vaginally born infants
- This deviation can be prevented by fecal microbiota transplantation from the mother (T3 feces; collected 3 weeks prior to delivery)
- Transplanted cesarean section-born infants show normal fecal microbiota development at 1-3 months of age.

7 infants received FMT
(1 time 3.5mg = 10^6 - 10^7 bacteria)

1-3 Months Intestinal Microbiota

Korpela *et al.* Cell 2020

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Primo-colonization - window of opportunity

A: Germ-free colon

- INKT cell colonization
- Susceptibility to **Colitis** (blue arrow)
- Susceptibility to **Colitis** (red arrow)
- Healthy colon (green arrow)
- Healthy colon (green arrow)
- Susceptibility to **Colitis** (orange arrow)

B: Germ-free lung

- Susceptibility to **Asthma** (blue arrow)
- Susceptibility to **Asthma** (red arrow)
- Healthy lung (green arrow)
- Susceptibility to **Asthma** (green arrow)

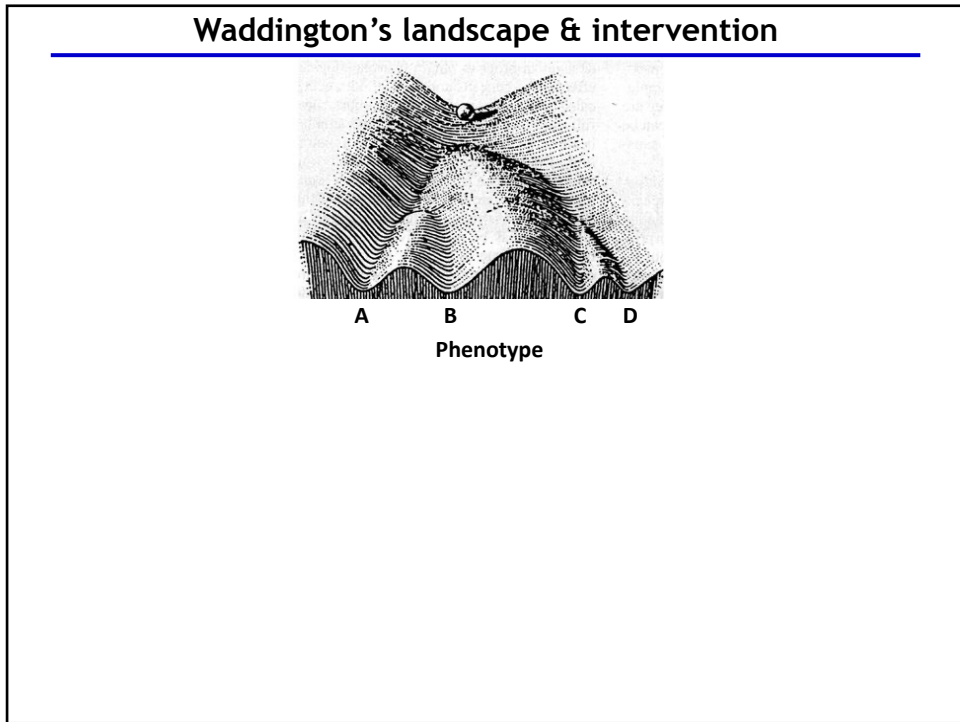
Birth Day 6-7 2 weeks Adult

Window of opportunity

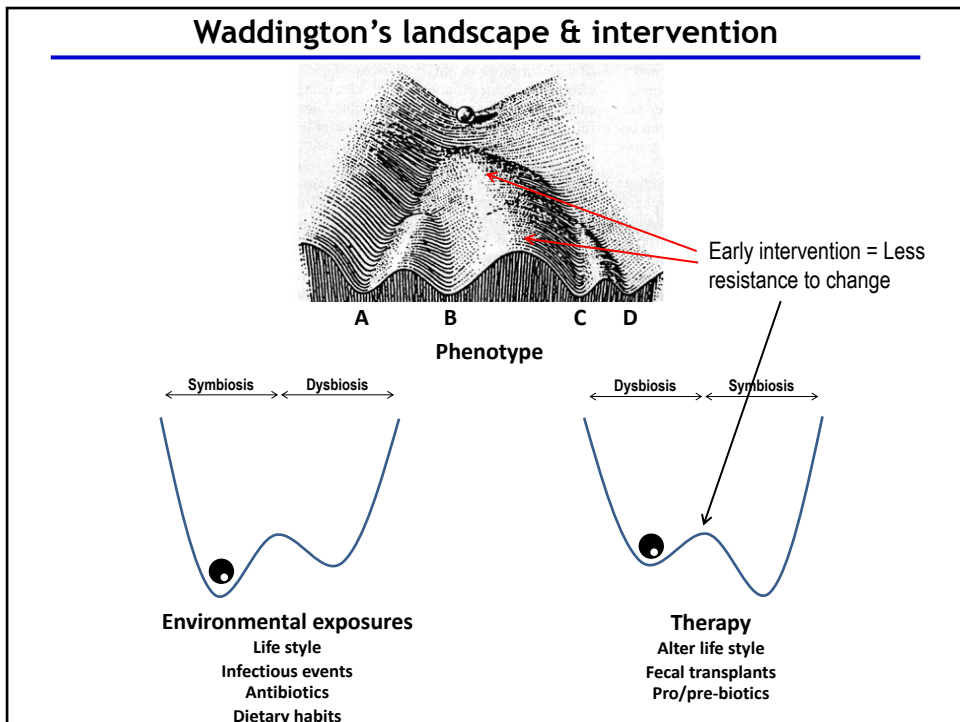
→ Germ-free → Standard microbiota → Bacteroides fragilis
→ Bacteroides fragilis strain lacking sphingolipids Conventionalization *B. frag* Monocolonization with Bacteroides fragilis *B. fragASPT* Monocolonization with Bacteroides fragilis mutant strain lacking sphingolipid

Gensollen *et al* Science 2016

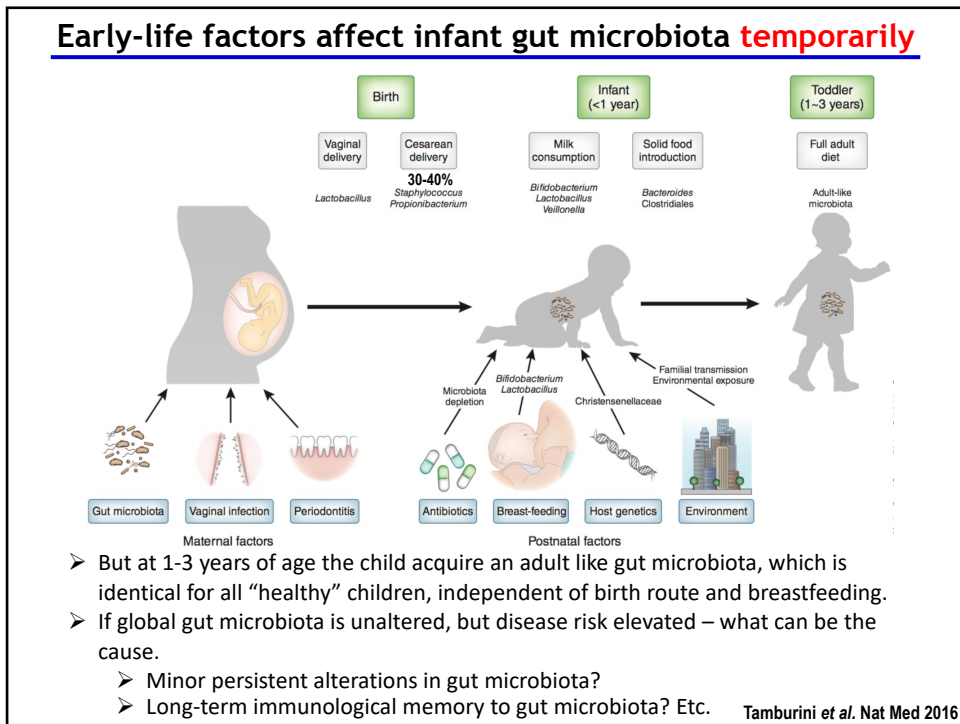
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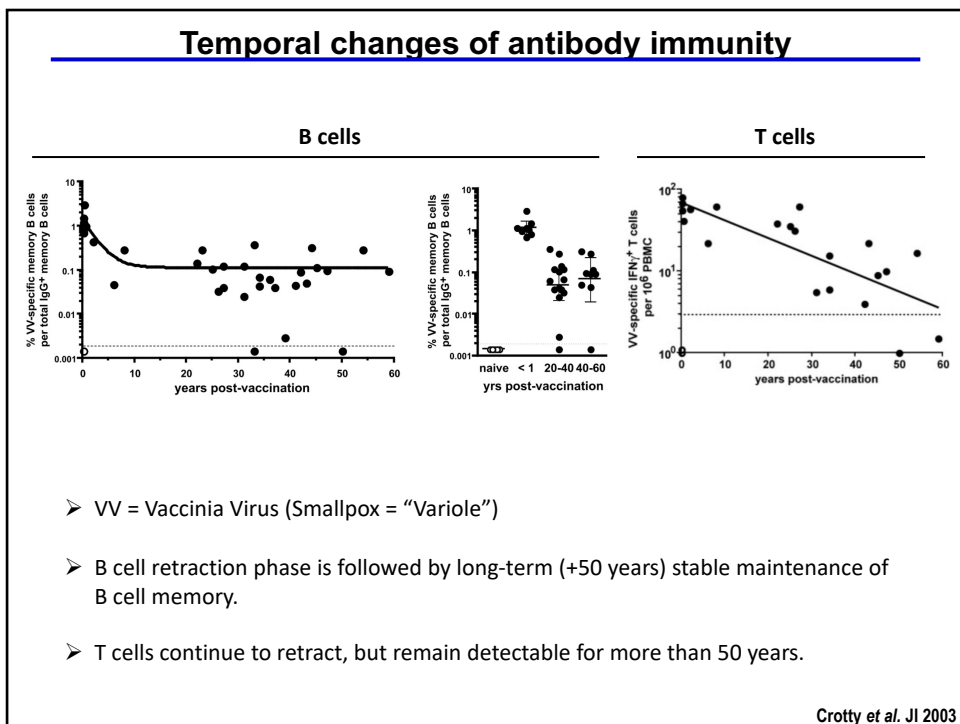
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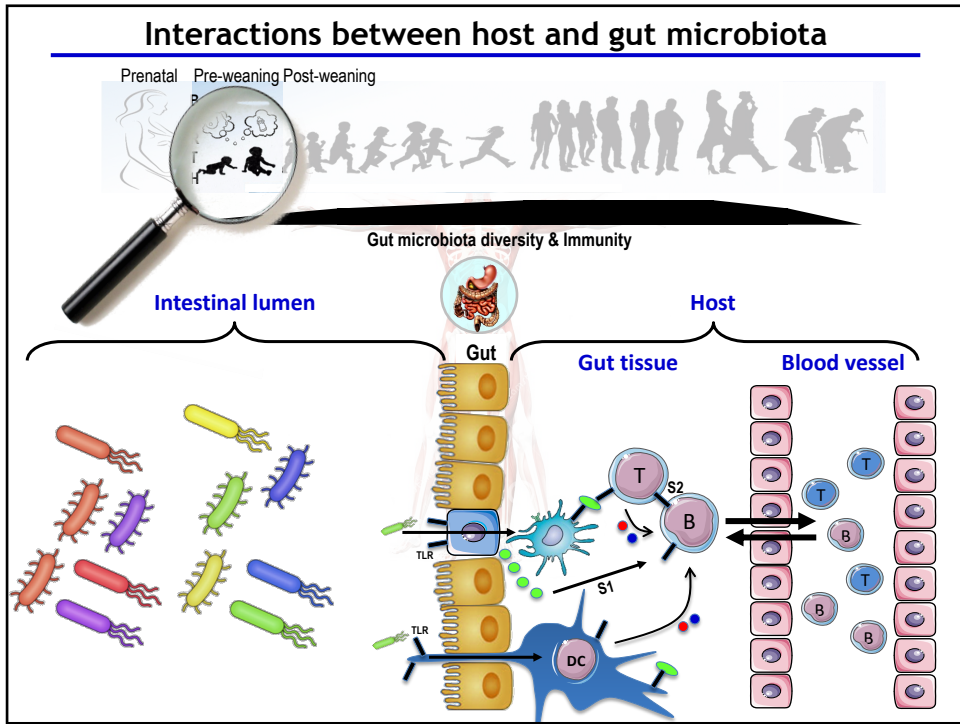
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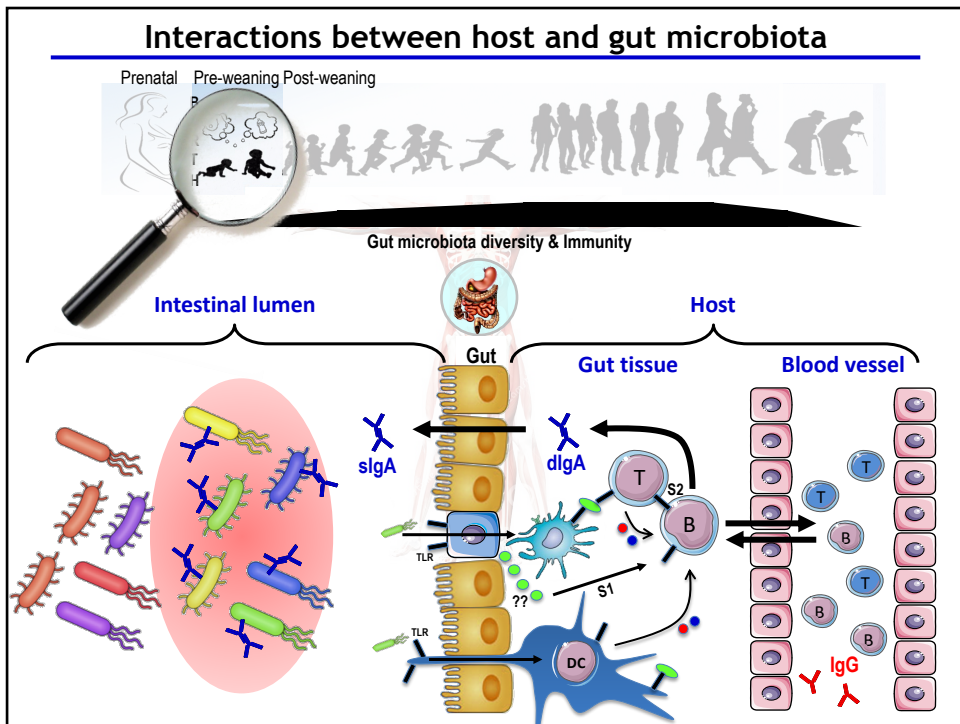
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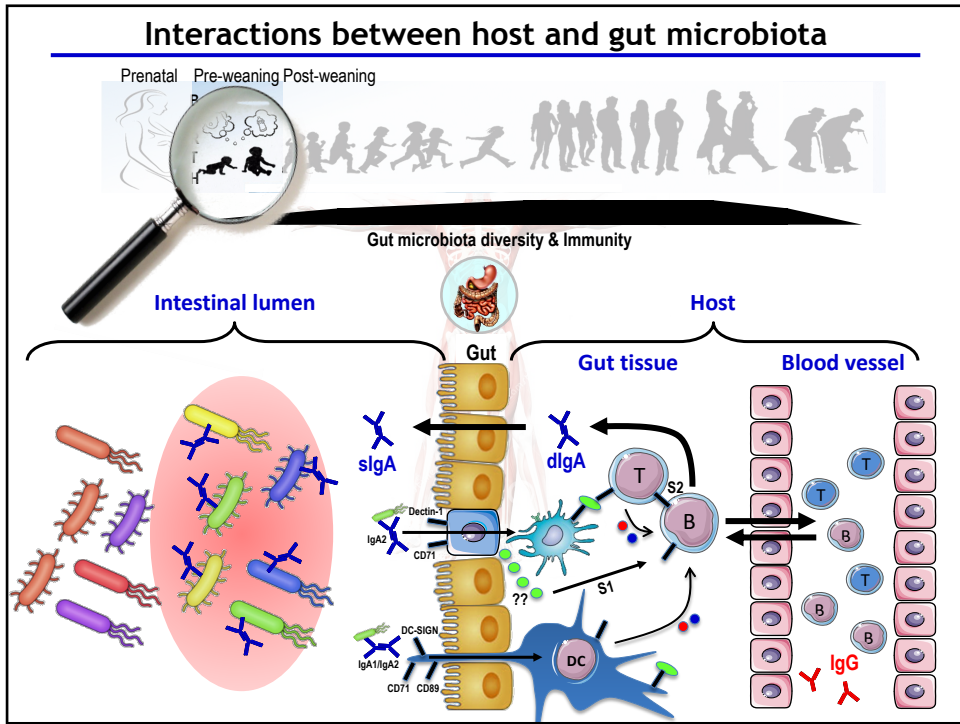
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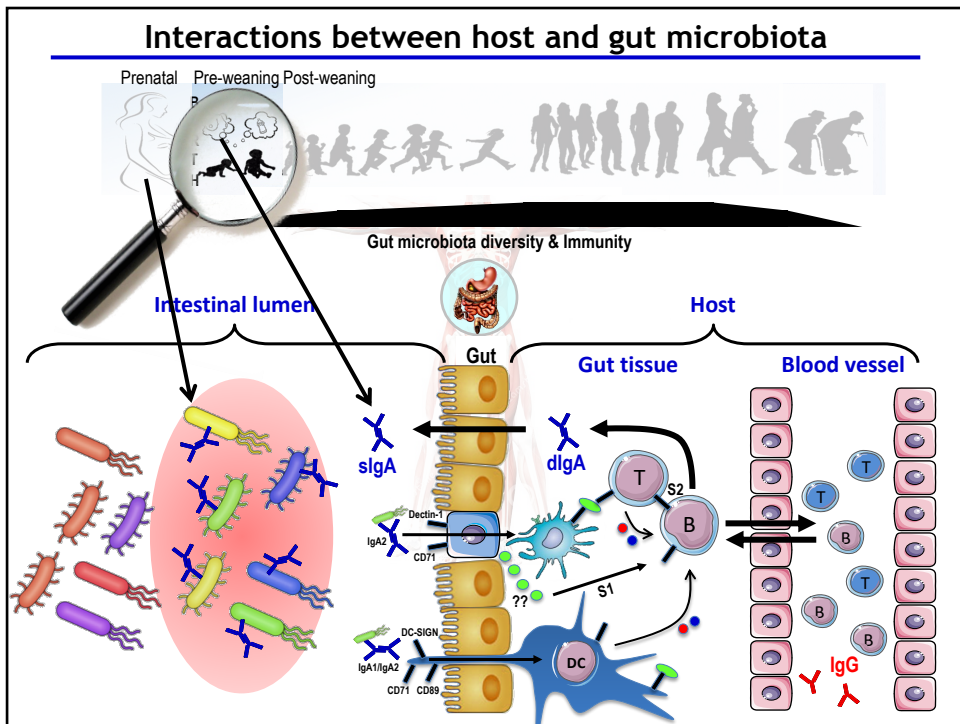
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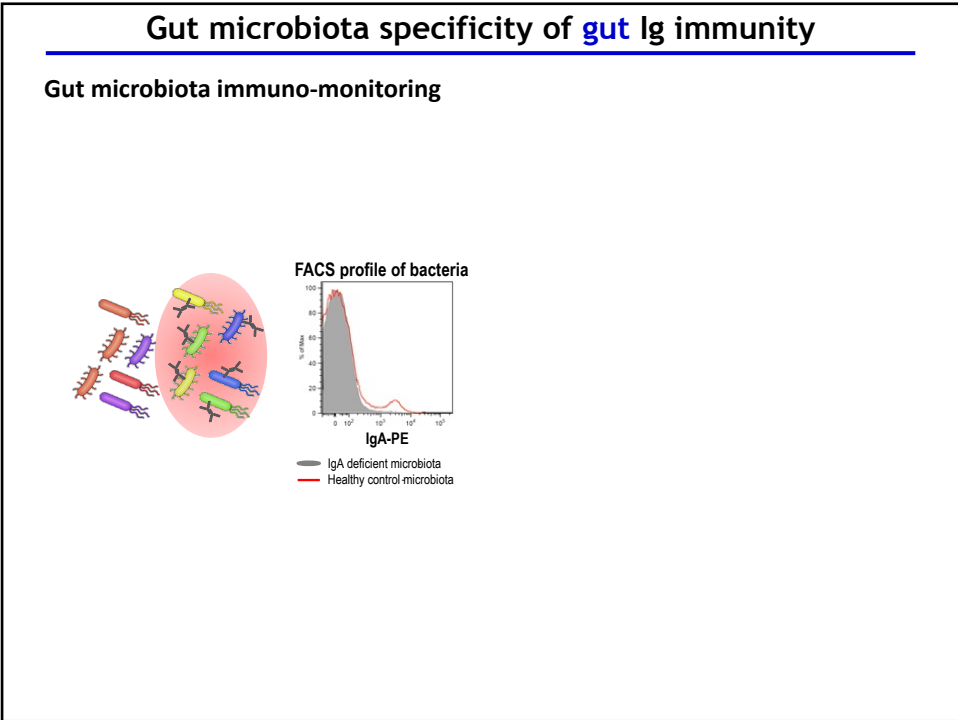
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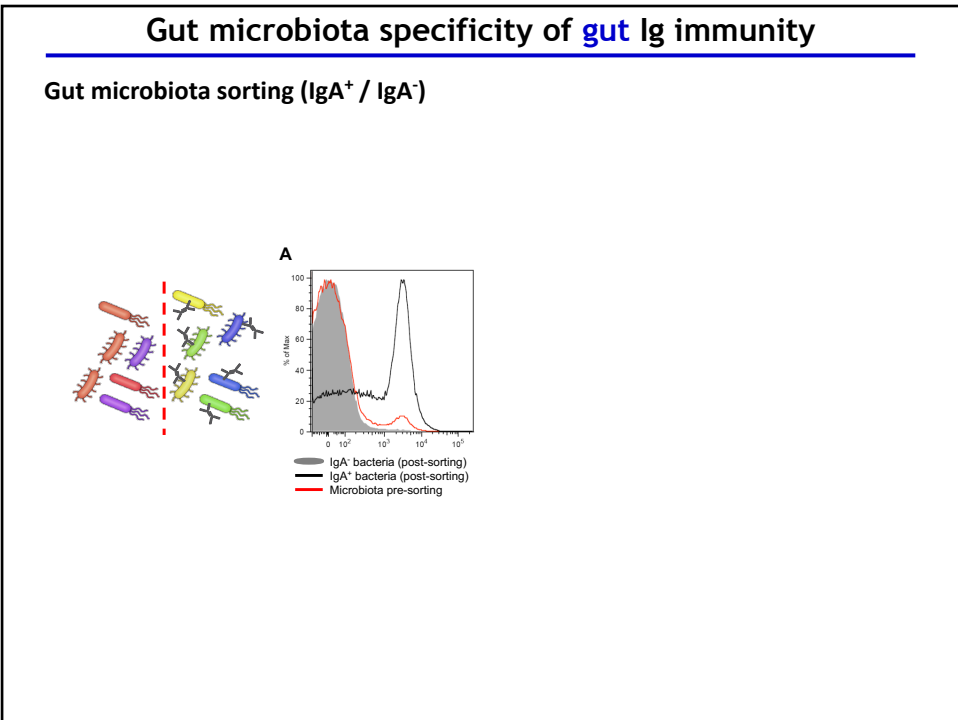
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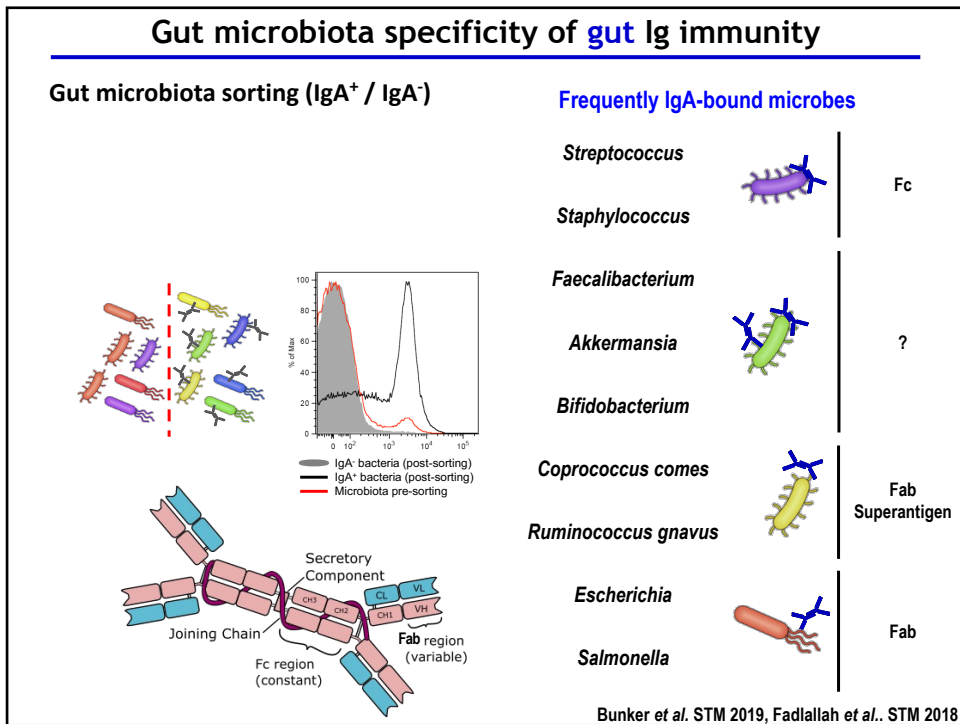
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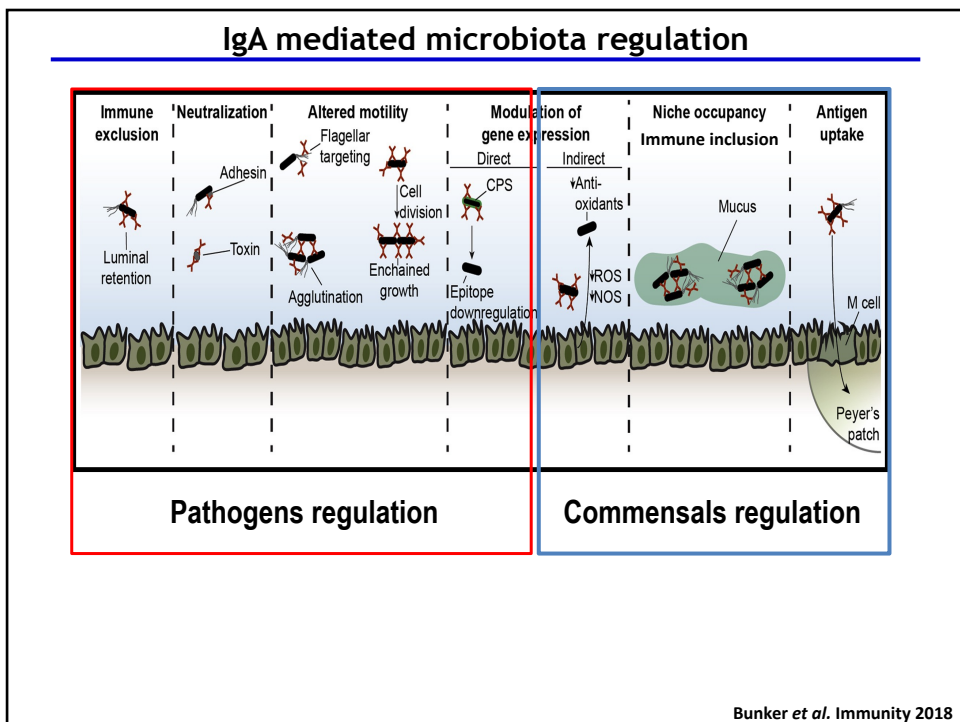
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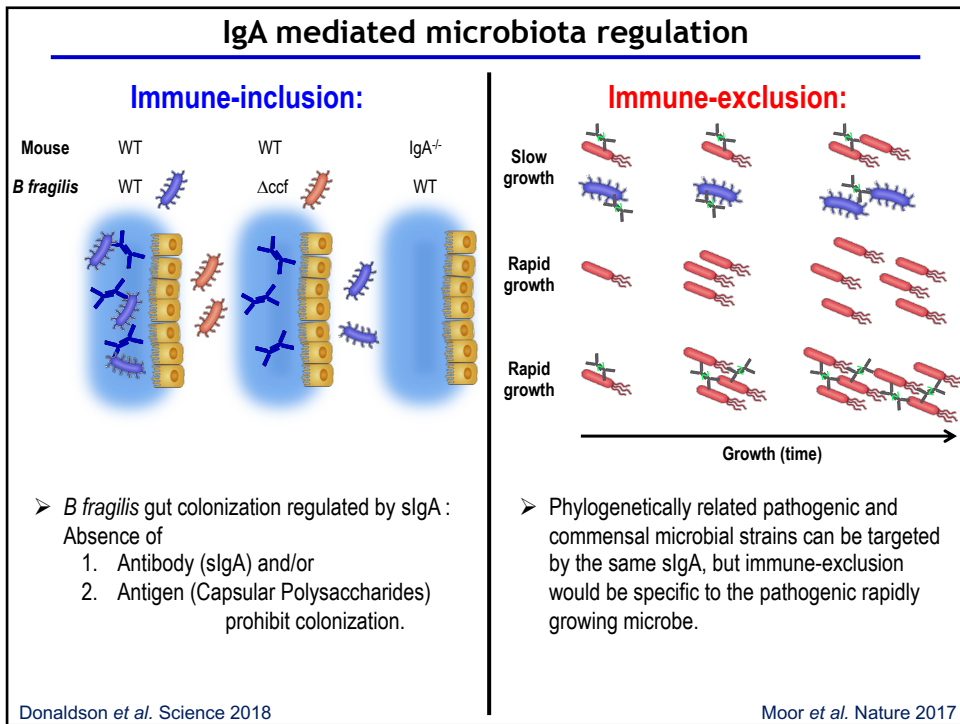
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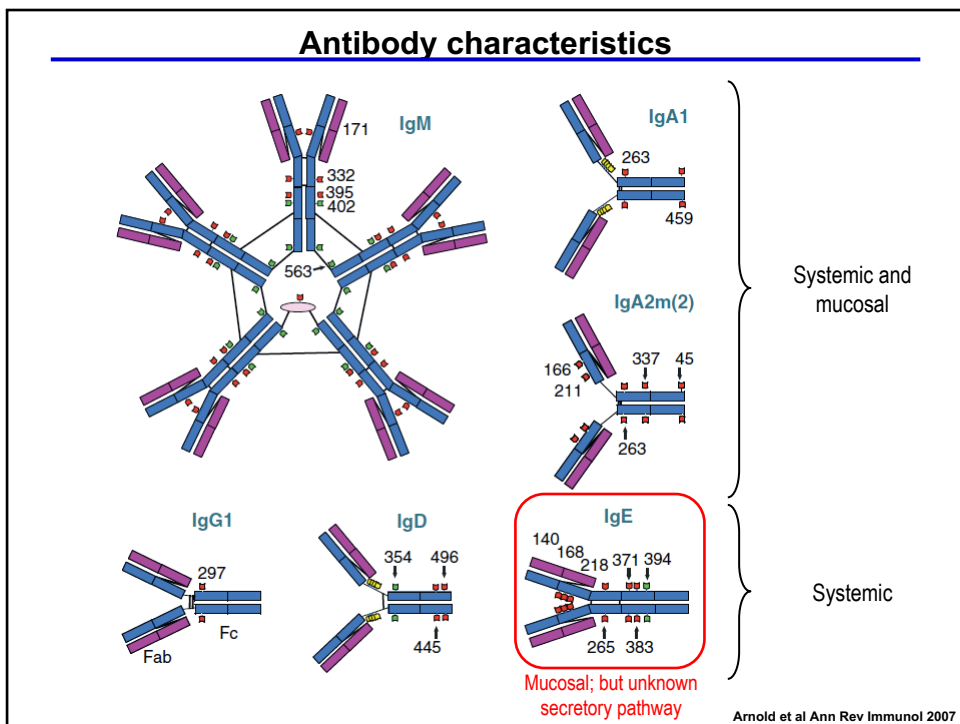
101



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Self/non-self versus Danger model in a historical perspective

- Burnet and Lederberg propose the antigen receptor (BCR and TCR)
- Antigen stimulation (signal 1) induces immunity including class switch and somatic hyper mutation.

Matzinger *et al.* Scan J Immunol 2003

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Self/non-self versus Danger model in a historical perspective

Signal 2: e.g. CD40/CD40L and cytokines

- **Problem:** BCR hypermutation may lead to autoreactive BCRs.
- **Solution:** Cohn add another cell: The T helper cell (only formally proven much later (Mosmann *et al.* 1986)).
- B cells internalize pathogen and present antigens to interact with specific Th cells, which validate that target is non-self.
- Signal 1 alone leads to clonal deletion (both self and non-self reactivity leads to signal 1).
- Signal 1 + 2 lead to activation (non-self rescued by T cells, with TCR which does not hyper mutate).

Matzinger *et al.* Scan J Immunol 2003

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Self/non-self versus Danger model in a historical perspective

Signal 2 co-stim: CD80/86 (B7), CD28, CTLA-4, CD40/CD40L

- **Problem:** We need a BCR independent manner to capture and present antigens to prime naïve T cells to become T helper cells.
- **Solution:** Lafferty and Cunningham propose that Th cells are primed and activated through APC antigen presentation and co-stimulation.
- Th cells are not constitutive active – feedback regulation upon antigen removal.
- Heavily criticized because APCs do not explain how the immune system distinguish between self and non/self (which BCR dependent antigen selection provided).

Matzinger et al. Scan J Immunol 2003

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Self/non-self versus Danger model in a historical perspective

- **Problem:** Contrary to B cells, APCs do not differentiate between self and non-self.
- **Solution:** Charlie Janeway proposes that APCs internalize and present non-self selectively through **pattern recognition receptors (PRR)**, which bind elements from foreign organisms, such as bacteria.
- This model inherently imply that APCs are not constitutively active, but require external stimuli through the PRR signalling pathway.
- Proposes explanation for why vaccines need an adjuvant.

Matzinger et al. Scan J Immunol 2003

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Self/non-self versus Danger model in a historical perspective

Problem

How to explain:

- Autoimmunity
- Non-reject of tumour with tumour antigen
- Why mothers don't reject the fetus.
- Why temporal gene-expression changes doesn't evoke immunity (e.g. breast milk).
- Why can we host billions of microbes?

1950

1960

1970

1980

1990

← 1996

2000

➤ **Solution:** Polly Matzinger proposes that APCs are activated through danger signals from any distressed tissue (uric acid, heparin sulfate, extracellular ATP and DNA/RNA).

➤ Advocate that we are a friendly host as long as our visitors are friendly too. Don't push the button first policy.

Matzinger et al. Scan J Immunol 2003

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Danger model and gut microbiota symbiosis

➤ The danger model would suggest that we do not respond to non-harmful bacteria colonizing our gut (Mutualism and Commensalism allowed).

Symbiosis
Living together (sym: with; bio: life)

Mutualism	Commensalism	Pathobiont	Parasitism
Both benefit	One benefit; one not harmed	Both benefit if regulated	One benefit; one harmed
Immune Ignorance (no immunity but barrier)		????	Cytotoxic Immunity

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Danger model and IgA responses

➤ The danger model would suggest that we do not respond to non-harmful bacteria colonizing our gut (Mutualism and Commensalism allowed).

Symbiosis
Living together (sym: with; bio: life)

Mutualism Both benefit	Commensalism One benefit; one not harmed	Pathobiont Both benefit if regulated	Parasitism One benefit; one harmed
} Immune Ignorance (no immunity but barrier)		} Immune Tolerance (e.g. IgA, Treg)	} Cytotoxic Immunity

➤ In reality microbes cannot be categorized discretely, but rather represents a continuum from Mutualism to parasitism.

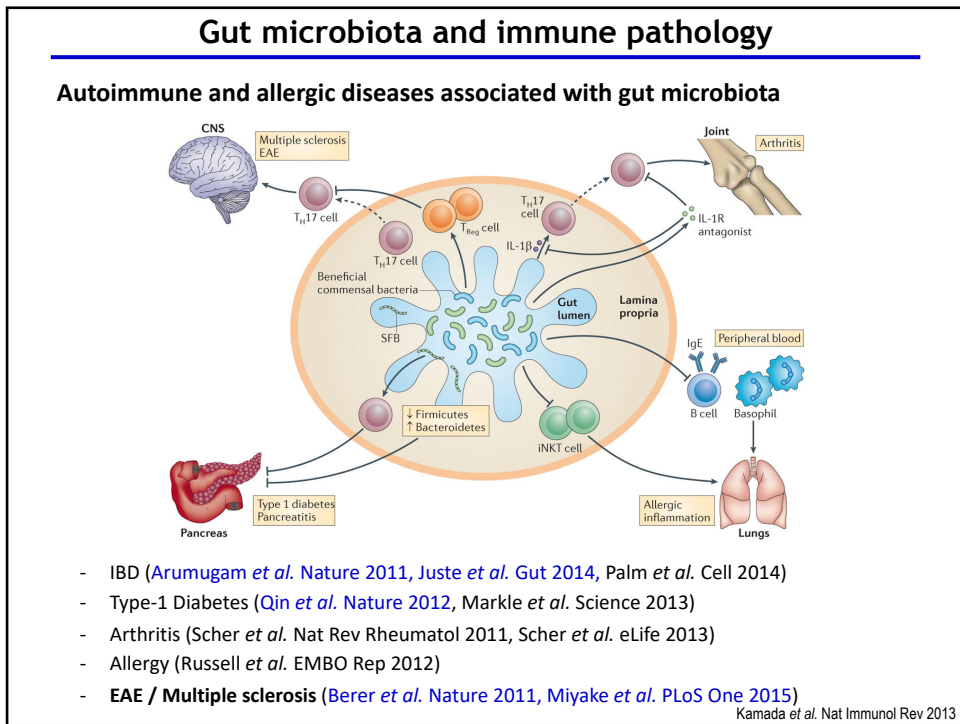
➤ How to retain tolerance to commensals, while pathogens are attacked?

➤ The range of host-microbe interactions evokes ignorance to non-harmful microbes, tolerogenic immunity to beneficial microbes (harmful if not regulated) and cytotoxic immunity to harmful microbes.

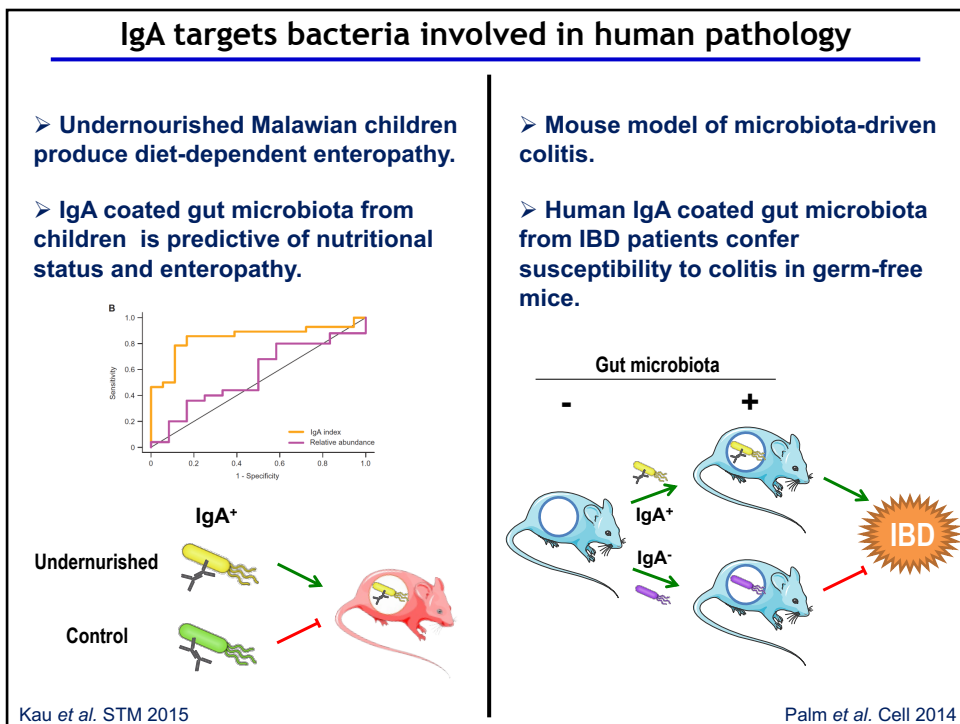
145

Examples of gut microbiota associations with pathology

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Dysbiosis - cause or consequence of disease?

- Genetic or environmental factors may lead to dysbiosis
- Dysbiosis may lead to disease
- Genetic or environmental factors may lead to disease irrespective of dysbiosis.
- Disease may lead to dysbiosis

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Study design defines the ability to determine causality

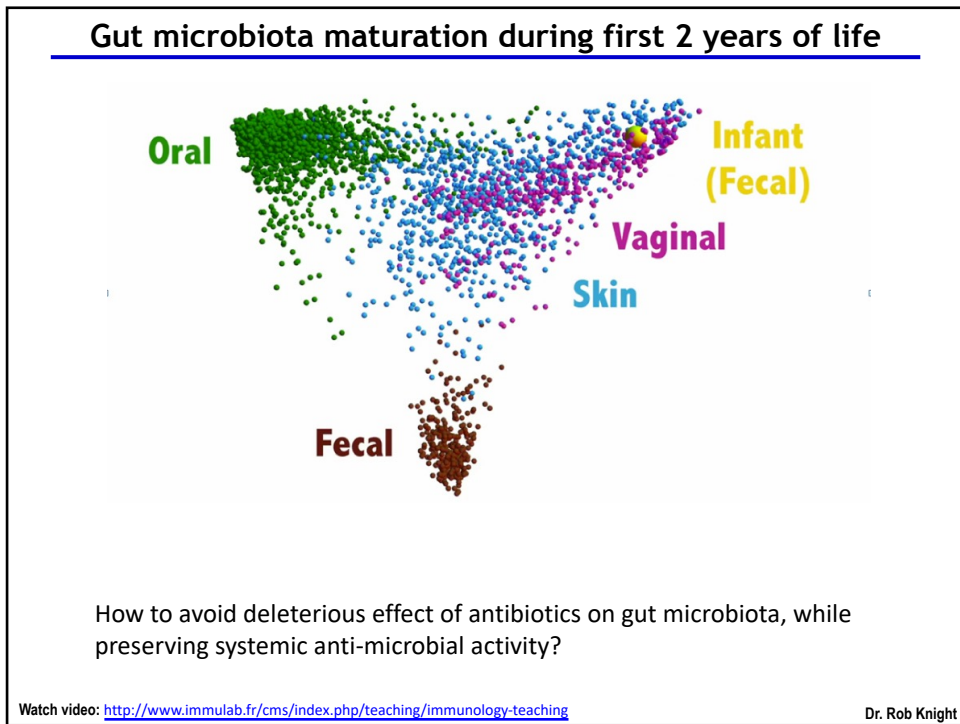
Evidence for causality

Interventional studies
Modulating microbiota composition alters health status.

Prognostic cohort studies
Discriminating microbiota patterns precede clinical outcome.

Cross-sectional case-control studies
Discriminating microbiota patterns associated with disease

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Avoid intestinal effect of antibiotics

- Antibiotics (yellow) administered orally for non-intestinal infections (dental, sinus, pulmonary etc.) are taken up primarily in the stomach. Remains pas through our lower GI tract.
- Activated coal (DAV132) released in the colon absorbs/inactivates 99% of antibiotics.

➤ DAV132 retains normal antibiotic plasma levels, but reduces fecal levels and protects gut microbiota from antibiotic induced dysbiosis.

Watch video: <http://www.immulab.fr/cms/index.php/teaching/immunology-teaching> De Gunzburg et al JCI 2017

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Take home message

- Gut microbiota influence host immunity (Tolerance versus inflammation)
- Gut microbiota is regulated by host immunity (innate and adaptive (e.g. IgA))
- **Altered lifestyle** influence our gut microbiota composition and is temporally (but maybe not causally) associated with a rapid increase in chronic inflammatory diseases, including allergy (since 1950 forward).
- **Hygiene theory:** Reduced exposure to microbes result in a skewed host immunity, which is insufficiently schooled to regulate inflammatory responses.
- **Save our microbiota:** Fecal (*C. difficile*) and Vaginal (C-section birth) microbiota transplantation, reduce antibiotics use (or use of new treatments, such as DAV132 co-therapy).
- **Save our immunity:** Probiotics (do not colonize), helminths (worms), immune therapy (allergy), promote breast feeding.